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Recent Genetic studies of Pacific jack mackerel *Trachurus murphyi* in
Russia

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In 2007 in Russia in the Belozersky Scientific-Research Institute of physico-chemical biology the Moscow State University initiated preliminary genetic research of Pacific jack mackerel. Samples were collected onboard the Dutch fishing vessel “Jan Maria”. I’d like to take this opportunity to express my thanks to Dr. Corten and the shipowner for the rendered opportunity for taking of these genetic samples. Since all the samples were taken at the comparatively small area not far from the EEZ of Chile between 37°48 and 38°28 S and 80°49 and 84°28 W, the undertaken activities were of preliminary nature and their objective was to elaborate methodical aspects for search of genetic polymorphous markers of population level. The method of restricted fragments length polymorphism (RFLP) mtDNA was applied in the research. The branded NucleoSpin®Tissue (MACHEREY-NAGEL) set was used for extraction of Pacific jack mackerel’s DNA. The DNA was extracted out of 50 samples of liver (Fig. 1). For RFLP analysis variable sectors of mtDNA-cytochromes b (1140 n.p.) and D-loop (800 n.p.) were chosen.

The following primers' sequences were used for amplification of these fragments:

Cytochrome b (Cardenas et al., 2005)

tRNA-Glu-CTB-F: 5`-ATG GCA AAT CTC CGT AAA ACC C-3`

t-RNA-Thr- CTB-R: 5`-AGG CTC ATC CGA GCA TTT TA-3`

D-loop (Poulin et al., 2004)

(tRNA-Pro)-tRNA T1-DI-F: 5`-CAG AAA AAG GAG ACT CTA ACT CCT AAA-3`

(tRNA-Phe)-tRNA T2-DI-R: 5`-TGC TTG CGG GGC TTT CTA-3`

Primers' sequence (tRNA-Pro)-tRNA T1-DI-F was changed as per GenBank:

5`-CAG AAA AAG GAG ACT CTA ACT CCT-3`

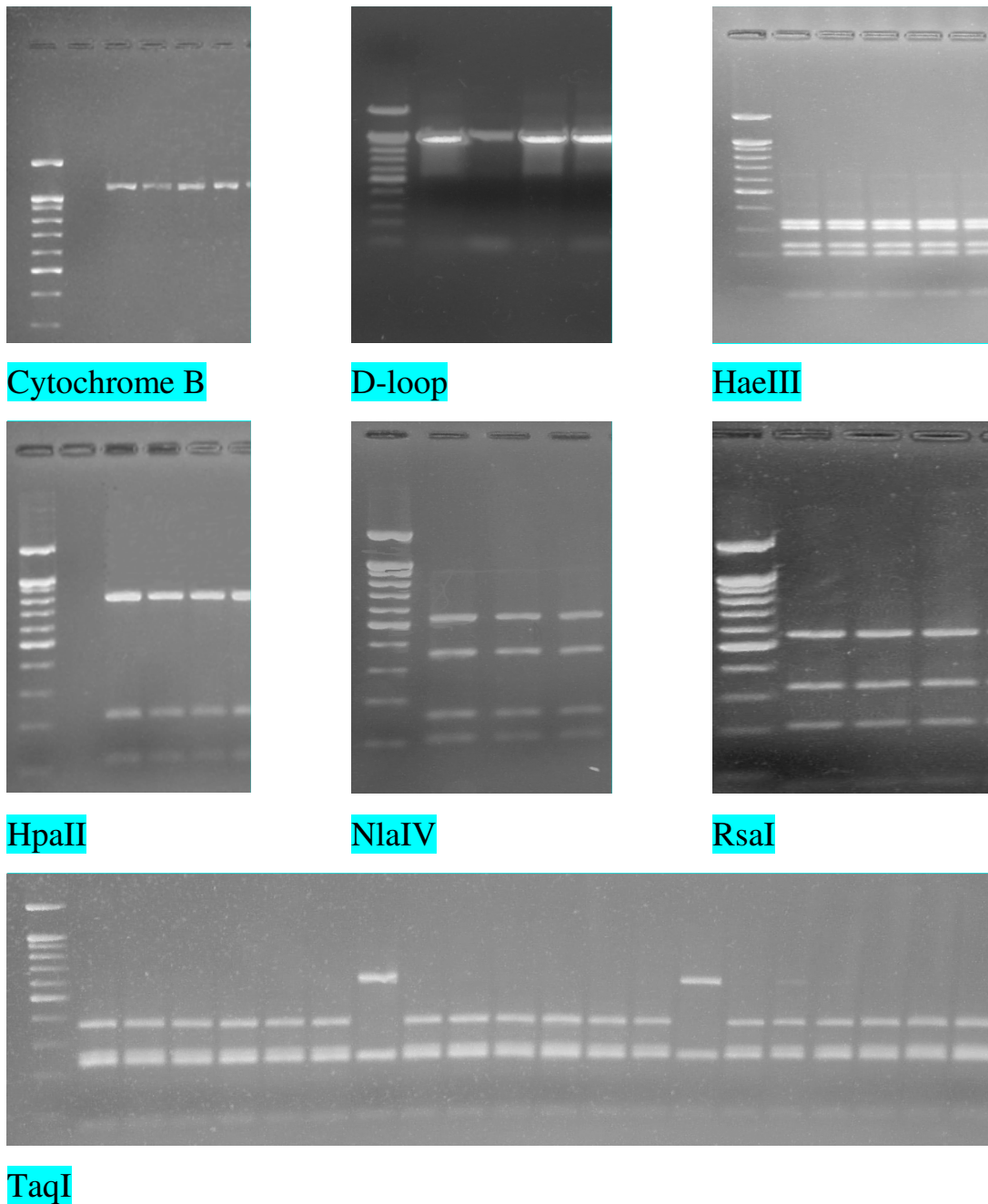
Cytochrome b and D-loop were amplified in the following conditions: initial denaturation at 95°C - 5 min., then 30 cycles consisting of denaturation at 94°C 30 sec, primers' annealing (e'niling) at 56°C - 45 sec, elongations at 72°C - 45 sec and the final elongation at 72°C - 10 min. Amplification of D-loop was carried out in the same conditions except for the elongation stage of 30 cycles: at 72°C - 90 sec. Amplification of mtDNA fragments was conducted in 25 µl of reaction mixture consisting of 2.5 µl of 10X buffer (Dialat Ltd.), 2 units of Taq-polymerase (Dialat Ltd.), 2.5 mM MgCl₂ (Dialat Ltd), 0.2 mM dNTPs (Dialat Ltd), 10 pM of each primer and 5-10 ng of DNA. Amplification of D-loop was carried out with the same volume of reaction mixture with increase of MgCl₂ concentration to 3.2 mM. (Fig. 2) The amplified fragments of mtDNA were studied for polymorphism of restricted fragments. Restricted endonucleases (restrictases) were used in this work.

Characteristics of the used restrictases.

Name of restrictase	T of restriction, °C	Recognition site	Restricted fragments, n.p.	Polymorphism of fragments' lengths
HaeIII (BsuRI)	37	GG^ CC	320+300+220+200+ 100	No
HpaII (MspI)	37	C^C GG	820+210+110	No
NlaIV (Psp N41)	37	GGN ^NCC	540+350+150+100	No
RsaI	37	GT^ AC	540+310+200+90	No
TaqI	60	T^C GA	370+240+220+220+ 90 610+220+220+90	Yes

Restriction of amplified fragments of mtDNA took place in 20 µl of reaction mixture, consisting of 2 µl of 10X buffer, 5 µl of the obtained amplicate and 1.5 µl of the appropriate restrictases. The restriction was carried out at optimal temperatures within 2 hours. Determining of lengths of amplified and restricted fragments was conducted in 2% agarous gel with 1XTAE buffer using 100 bp+1.5 Kb DNA ladder (NPO SibEnzim) marker with further colouring by ethidium bromide (0.5 mg/ml).

(Fig. 3) As a result of the performed research: DNA was extracted out of 50 samples of Pacific jack mackerel tissue. Variable fragments of mtDNA cytochrome b and D-loop have been amplified. RFLP analysis of cytochrome b was carried out with use of HaeIII, HpaII, NlaIV, RsaI, TaqI restrictases. Polymorphism of restricted fragments was determined with use of TaqI.



Fragments of Cytochrome b obtained as a result of restriction by the restrictases signed under the figures

In conclusion I'd like to discuss some methodological aspects of future research activities. In the report prepared by Chile reference is made to the work of Ojeda and Poulin, 2002 regarding testing of 4 microsatellite markers for *T. murphyi*. Since until now there has been no information concerning availability of sequences of *T. murphyi* or *T. symmetricus* microsatellite fragments in the Genetics

Bank, it seems that the heterologous primers of related species, i.e. *Gnathonodon speciosus* or *Seriola dumerili* (Feng et al, 2005, Ohara et al, 2003, Babbucci et al, 2006), sequences of microsatellite loci of which are available in the Genetics Bank or published., were used while performing of this work.

At the end of 2007 the nucleotide sequences of some microsatellite loci of *Trachurus trachurus* and *Trachurus japonicus* appeared in the Genetics Bank, however all of them are dinucleotide (registration numbers are EF 109781-EF 109801 for *T.japonicus* and EU 748-EU 751 – for *T.trachurus*).

For Mediterranean scad and Atlantic jack (blue scad) differentiation there were used four specially developed dinucleotide microsatellite markers (the same as the above mentioned of the Genetics Bank), which proved to be low effective. In the *Fisheries Research* publication (Kasapidis & Magoulas, 2008) there were given characteristics of these loci, including polymorphism – 40-42 and even 65 alleles. Samplings of up to 1000 specimens are necessary for proper use of such markers. There is a probability that inefficiency of microsatellite analysis in this case was caused by non-optimal and not quite sufficient set of microsatellites. This is also pointed out by the authors of that paper, proposing to consider the results of the work to be preliminary.

Tetranucleotide microsatellite sequences are more effective for the purposes of population analysis. However, development of such primers is considerably more expensive compared with the same for dinucleotides.

(Fig. 4) The main conclusions which can be made as a result of studying the existing literature on identification of Horse mackerel stocks are as follows:

The main literature conclusions

1. Mitochondrial DNA is not characterized by sufficient polymorphism in respect of intraspecific jack mackerel's groups.

2. Allozyme analysis is quite perspective, however requires selection of enzymatic systems with more moderate polymorphism in respect of Jack mackerel's populations.

3. At present developing of specific microsatellite markers, the number of nucleotides of which equals to 4 and over, is the most perspective tendency of jack mackerel's population genetic research. However, such research activities require significant temporal and financial inputs.

4. The Single Strend Conformational Polymorphism - analysis is perspective for separation of jack mackerel's populations

(Fig. 5)

1. Abaunza P., Gordo L., Karlou-Riga C., Murta A., Eltink A.T.G.W., García-Santamaría M.T., Zimmermann C., Hammer C., Lucio P., Iversen S.A., Molloy J., Gallo E., 2003a. Growth and reproduction of horse mackerel, *Trachurus trachurus* (Carangidae). *Reviews in Fish Biology and Fisheries* **13**:27–61.

2. Abaunza, P., Campbell, N., Cimmaruta, R., Comesaña, S. Dahle, G., Gallo, E., Gordo, L., Iversen, S., MacKenzie, K., Magoulas, A., Mattiucci, S., Molloy, J., Murta, A., Nascetti, G., Pinto, A.L., Quinta, R., Ramos, P., Ruggi, A.,

Sanjuan, A., Santamaría M.T., Santos, A.T., Stransky, C., Terzoglou, V., Zimmermann, C., 2003b. Final Report of the EU funded project HOM SIR: “A multidisciplinary approach using genetic markers and biological tags in horse mackerel (*Trachurus trachurus*) stock structure analysis”. Code: QLK5-Ct1999-01438.

3. Abaunza P., Murta A, Mattiucci S., Cimmaruta R., Nascetti G., Magoulas A., Sanjuan A, Comesaña S., MacKenzie K., Molloy J., Santos A.T, Iversen S, G. Dahle, Gordo L., Stransky C., Zimmermann C., Santamaria M.T., Ramos P., Quinta R., Pinto A.L., Ruggi A, Campbell N., 2004. Stock discrimination of horse mackerel (*Trachurus trachurus*) in the Northeast Atlantic and Mediterranean Sea: integrating the results from different stock identification approaches. ICES CM 2004/EE:19 “Not to be cited without prior reference to the author”

4. Abaunza, P., Murta, A.G., Campbell, N., Cimmaruta, R., Comesana, A.S., Dahle, G., Gallo, E., Garcya Santamaría, M.T., Gordo, L.S., Iversen, S.A., MacKenzie, K., Magoulas, A., Mattiucci, S., Molloy, J., Nascetti, G., Pinto, A.L., Quinta, R., Ramos, P., Ruggi, A., Sanjuan, A., Santos, A.T., Stransky, C., Zimmermann, C., 2008 a. Considerations on sampling strategies for an holistic approach to stock identification: the example of the HOM SIR project. Fish. Res., v.89, p. 104-113

5. Abaunza P., Murta A.G., Campbell N., Cimmaruta R., Comesana A.S., Dahle G., Garcya Santamaría M.T., Gordo L.S., Iversen S.A., MacKenzie K., Magoulas A., Mattiucci S. , Molloy J., Nascetti G., Pinto A.L, Quinta R., Ramos P., Sanjuan A., Santos A.T. , Stransky C., Zimmermann C., 2008 b. Stock identity of horse mackerel (*Trachurus trachurus*) in the Northeast Atlantic and Mediterranean Sea: Integrating the results from different stock identification approaches. Fisheries Res., v. 89, p. 196–209.

6. Babbucci M., Zane L., Andaloro F., Patarnello T. 2006. Isolation and characterization of microsatellite loci from yellowtail *Seriola dumerilii* (Perciformes: Carangidae). *Molecular Ecology Notes*, V 6, N 4, p. 1126-1128.

7. Cardenas L., Hernandez C.E., Poulin E., Magoulas A., Kornfield I., Ojeda F.P. 2005. Origin, diversification and historical biogeography of the genus *Trachurus* (Perciformes: Carangidae) // *Molecular Phylogenetics and Evolution*. V. 35. 496-507 p.

8. Cimmaruta R., P. Bondanelli, A. Ruggi, G. Nascetti, 2008. Genetic structure and temporal stability in the horse mackerel (*Trachurus trachurus*). *Fish. Res.*, v. 89, p. 114–121

9. Comesana A. S., Martýnez-Areal M. T., Sanjuan A., 2008. Genetic variation in the mitochondrial DNA control region among horse mackerel (*Trachurus trachurus*) from the Atlantic and Mediterranean areas. *Fish. Res.*, v.89, p. 122-131.

10. Eschmeyer W. N (ed.), 2003. Catalog of fishes. Updated database version of March 2003. Catalog databases as made available to FishBase in March 2003. World Wide Web electronic publication. Available from: www.fishbase.org.

11. Feng F., Lo L.C., Lin Z., Zhu Y., Yue G. H., 2005. Isolation and haracterization of microsatellites in a marine food fish species, golden trevally *Gnathanodon speciosus*. *Mol. Ecol.Notes*, v. 5, p. 760-761.

12. Galleguillos R., Torres R., 1988. Identificación de unidades poblaconales pelagicas. IFOP. PUC. Talcahuana.

13. Johns G. C., Avise J. C., 1998. A comparative summary of genetic distances in the vertebrates from the mitochondrial Cytochrome b gene. *Mol. Biol. Evol.*, v.15, p. 1481-1490.

14. Karaiskou N., Triantafyllidis A., Triantafyllidis C., 2004. Shallow genetic structure of three species of the genus *Trachurus* in European waters. *Mar. Ecol. Prog. Ser.* v.281, p. 193–205.

15. Kasapidis P., Magoulas A., 2008. Development and application of microsatellite markers to address the population structure of the horse mackerel *Trachurus trachurus*. *Fisheries Res.* v. 89, p. 132–135

16. Naish K.W., 1990. The stock identification of the Cape Horse mackerel, *Trachurus trachurus capensis* (Pisces: Carangidae). Unpublished MSc. thesis, Rhodes University, South Africa.

17. Nekrasov V.V., 1994. Mackerel of the World Ocean (genus *Trachurus*) VNIRO, Moscow, pp. 1-228.

18. Ojeda P., Poulin E., 2002. Identificación de Unidades de stock del jurel (*Trachurus murphyi* Nichols) en el Pacífico Sudoriental mediante Análisis de marcadores Moleculares. Inform final PUC.

19. Ohara E., Nishimura T., Sakamoto T., Nagakura Y., Mushiake K. and Okamoto N., 2003. Isolation and characterization of microsatellite loci from yellowtail *Seriola quinqueradiata* and cross-species amplification within the genus *Seriola*. *Mol. Ecol. Notes* 3, 390-391

20. Poulin E., Cardenas L., Hernandez C. E., Kornfield I., Ojeda F. P., 2004. Resolution of the taxonomic status of Chilean and Californian Jack mackerels using mitochondrial DNA sequences. *J. Fish Biol.*, 65, 1160-1164.

21.. Reed D. L, Carpenter K. E, deGravelle M. J., 2002. Molecular systematics of the Jacks (Perciformes: Carangidae) based on mitochondrial cytochrome *b* sequences using parsimony, likelihood, and Bayesian approaches. *Mol. Phyl. Evol.*, v. 23, p. 513-524.

22. Smolenski A. J., Ovenden J. R., White R. W. G. 1994. Preliminary investigation of mitochondrial DNA variation in Jack Mackerel (*Trachurus declivis*, Carangidae) from South-eastern Australian waters. *Aust. J. Mar. Freshwater Res.* ,v. 45: p. 495-505.

23. Takashima Y., Morita T., Yamashita M. 2006. Complete mitochondrial DNA sequence of Atlantic horse mackerel *Trachurus trachurus* and molecular identification of two commercially important species *T. trachurus* and *T. japonicus* using PCR-RFLP. *Fisheries science* 72: 1054-1065.

(Fig. 6)

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